# Novel structural models of the Creatine transporter (SLC6A8) rationalize its structural determinants of binding

#### Introduction

Creatine is a crucial metabolite that plays a fundamental role in ATP homeostasis in tissues with highenergy demands. The creatine transporter (CreaT, SLC6A8) belongs to the solute carrier 6 (SLC6) transporters family, and more particularly to the GABA transporters (GATs) subfamily. Understanding the molecular determinants of specificity within the SLC6 transporters in general, and the GATs in particular is very challenging due to the high similarity of these proteins.

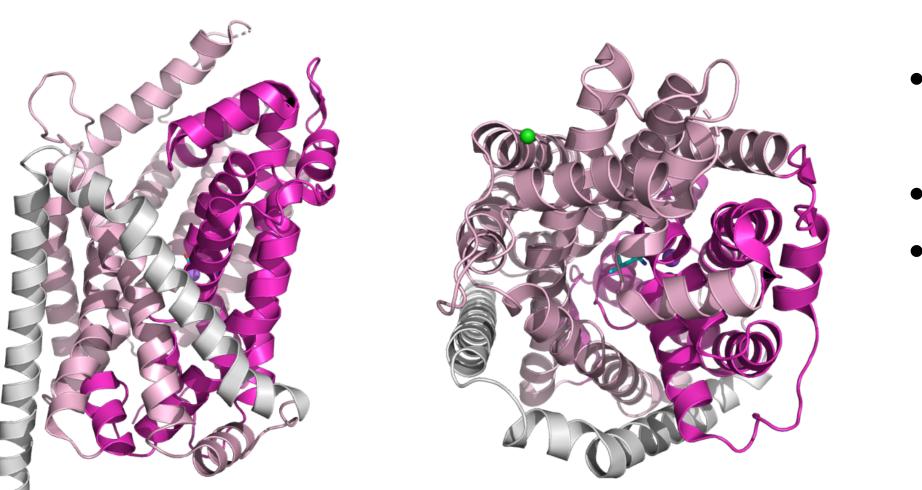
# **Goals and methods**

computational methods.

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- Building reliable homology models using two templates :  $\Rightarrow$  the human serotonin transporter (hSERT)<sup>1</sup>– outward open conformation
- $\Rightarrow$  the prokaryotic leucine transporter (LeuT)<sup>2</sup> occluded conformation

# **Templates : hSERT and LeuT**



**Three-dimensional structure of LeuT,** (PDB ID 2A65<sup>2</sup>) from side and top view respectively.

### **Sequence alignments**

LeuT	396	- L	N K	SL	D	ЕМ	D	F	W	A	3	- 1	гт	G	v	v	F 1	F	3 1	с 7	r e			I	F	F	w	I
SLC6A4 (SERT)	484																											
SLC6A3 (DAT)	467																											
SLC6A2 (NET)	464																											
SLC6A6 (TauT)	450	G G	мч	VF	Q	LF	D	Y	Y	A	A	s g	v	С	L	L	W	V J	<b>A</b> 1	FI	? E	C C	F	' V	I	A	W	I
<pre>SLC6A8_(CreaT)</pre>	465	G G	мч	VF	Q	L F	D	Y	Y	s i	A	s g	T	т	L	L	wş	2	<b>A</b> ]	FW	I	I C	V	7 V	v	A	W	v
<pre>SLC6A1_(GAT1)</pre>	442	G G	ΙY	VF	ĸ	ЬF	D	Y	Y	s i	A	s G	м	s	L	г	F ]	Г,	V	FI	? I		. v	7 5	I	S	W	F
<pre>SLC6A13_(GAT2)</pre>	438	G G	мч	VF	Q	L F	D	Y	Y	A	A	s G	M	С	L	L	F	V J	A	IF	? I	1 8	S I	C	V	A	W	v
<pre>SLC6A11_(GAT3)</pre>	458	G G	мч	IF	Q	ЬF	D	S	Y	A	A	s G	M	С	L	г	F	V J	A	IF	? I	I C		IC	I	G	W	v
<pre>SLC6A12_(BGT1)</pre>	443	G G	мч	IF	Q	LΓ	D	Y	Y	A	S	s g	1	С	L	L	FI	ь	S 1	LΒ	F	I V	7 V	7 C	I	s	W	v
<pre>SLC6A9_(GlyT1)</pre>	519	ΑG	ΙY	WL	г	LМ	D	N	Y	A	A	– S	5 F	s	L	v	V	I	s o	2	IM		V.	7 A	I	М	Y	I
<pre>SLC6A5_(GlyT2)</pre>	567	G G	ΙY	ΜF	Q	ъv	D	т	Y	A	A	- 5	S Y	A	г	v	I	I	A	IF	? E	1	J V	7 G	I	s	Y	v
SLC6A14	469	ΑG	ΙY	wν	н	LІ	D	H	F	<u>c</u> 1	A	–G	W	G	I	L	I	A	A	II	E	I	J V	7 <mark>G</mark>	I	I	W	I
SLC6A7_(PROT)	445	G G	мч	WL	v	LГ	D	D	Y	s i	A	- S	5 F	G	г	М	v١	v	V	I	<b>r</b> ?	٢C	: I	A	v	т	R	v
SLC6A20	452	ΑG	ΝY	WF	D	IF	N	D	Y	A	A	- 1	гL	s	L	г	г	I	V I	с 1	7 E		<b>c</b> :	IA	v	С	Y	v
SLC6A15 (BOAT2)	516	SG	ΝY	FV	т	ΜF	D	D	Y	s i	A	- 1	r I	Р	L	L	ľ	v	V	II	I	I N	r :	IA	v	С	F	v
<pre>SLC6A17_(NTT4)</pre>	515	SG	ΝY	FV	т	ΜF	D	D	Y	S 2	A	- 1	гL	Р	г	т	г	I	V	II	E	ľ	r :	IA	v	A	W	I
<pre>SLC6A19_(B0AT1)</pre>	477	SG	QY	WL	s	LГ	D	s	Y	A	3	– s	3 1	Р	L	г	I	I	<b>A</b> 1	FC	: 1	M	E	' s	v	v	Y	v
<pre>SLC6A18_(B0AT3)</pre>	463	SG	ΝY	WΙ	Е	IF	D	N	F	A	A	– s	5 P	N	L	L	<b>M</b> 1	ь	<b>A</b> ]	FI	E	1	7 V	7 G	v	v	Y	v
<pre>SLC6A16_(NTT5)</pre>	523	SG	SY	FΙ	R	LГ	S	D	Y	W	I	– V	7 F	Р	I	I	v	v	v	VI	F		ГМ	A	v	S	W	А

The sequence alignments of the SLC6 family reveals an additional amino acid in TM10 of all the GATs subfamily, suggesting the presence of a  $\pi$ -helix.

1. Coleman, J. A., Green, E. M. & Gouaux, E. X-ray structures and mechanism of the human serotonin transporter. Nature 532, 334-339, doi:10.1038/nature17629 (2016). 2. Yamashita, A., Singh, S. K., Kawate, T., Jin, Y. & Gouaux, E. Crystal structure of a bacterial homologue of Na+/Cl--dependent neurotransmitter transporters. Nature 437, 215-223 (2005).

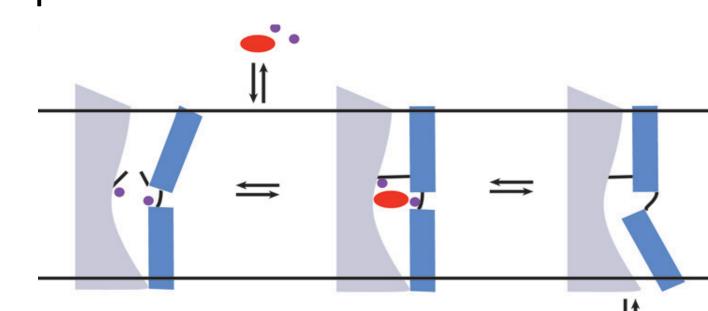
3. Colas, C., Ung, P. M. & Schlessinger, A. SLC Transporters: Structure, Function, and Drug Discovery MedChemComm 7(6):1069-1081 (2016) 4. Smith, R. H. B., Dar, A. C. & Schlessinger, A. PyVOL: a PyMOL plugin for visualization, comparison, and volume calculation of drug-binding sites. bioRxiv (2019).

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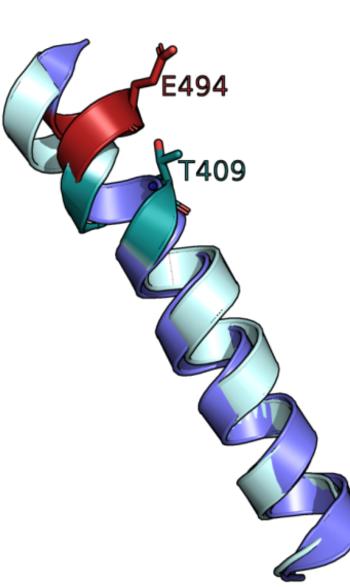
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Finding key structural features involved in binding selectivity for CreaT using structure-based

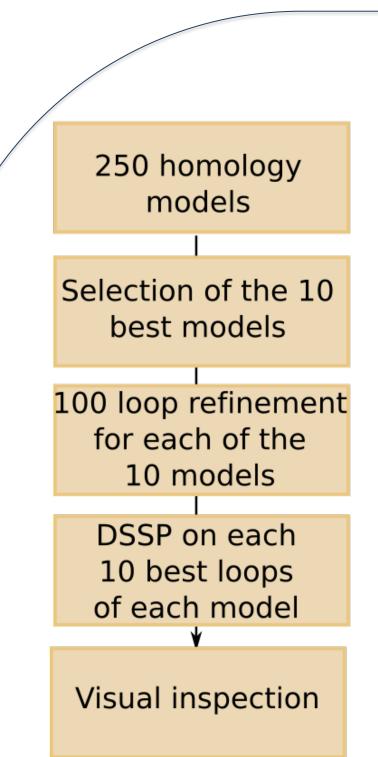
- LeuT and hSERT share 21% and 44% sequence identity with human CreaT respectively
- Similar"LeuT-fold"
- Use the gated-pore mechanism

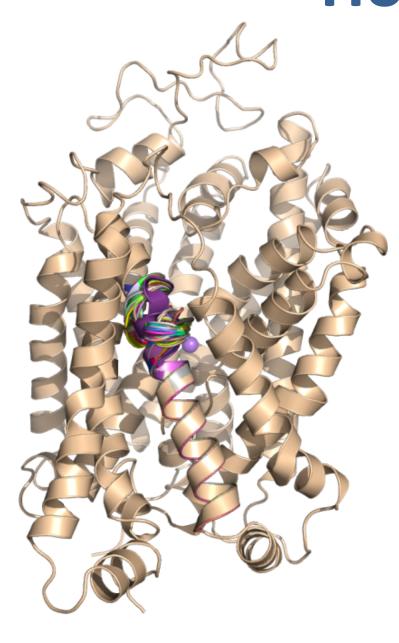


Gated-pore mechanism of transport<sup>3</sup>



 $\pi$ -helices identified distinct were positions in the templates LeuT (cyan) and SERT (red).





# Homology modeling

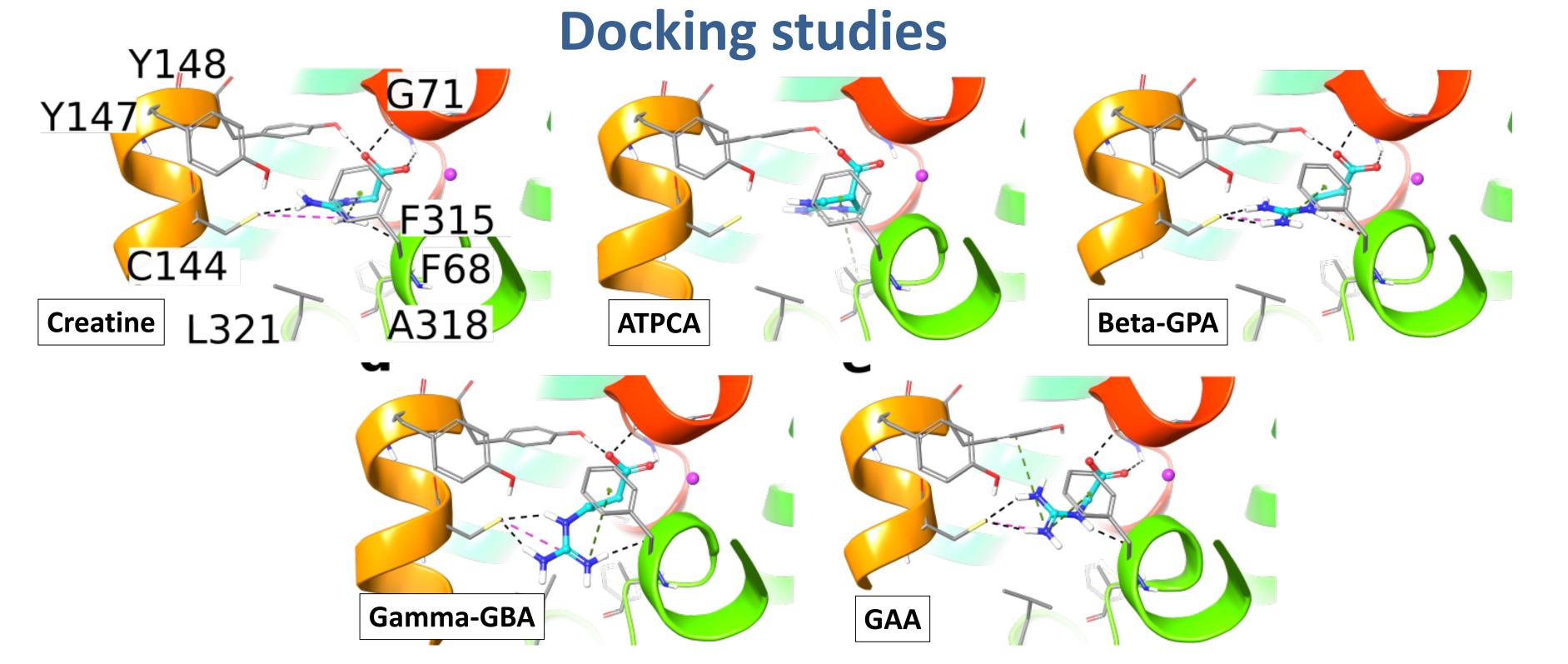
- occluded conformation
- TM10 insertion.

Main differences between the two conformations :

- a tilting of two broken helices TM1 and 6 on the extracellular side,
- a flipping of the conserved Tyrosine 148 (hydrophobic extracellular lid).

=> Significant increase of the binding site volume,<sup>4</sup> i.e. 285

Å<sup>3</sup> in the outward occluded conformation vs. 1293 Å<sup>3</sup> in the outward open conformation.



- Induced fit docking of known CreaT ligands in the occluded conformation to rationalize biological activities.
- An optimal length of carbon linker (4.5-5 Å) seems necessary between the guanidine and carboxylate groups to establish hydrogen bonds with respectively Cys 144 and Gly 71 and the Na<sup>+</sup>.

# Conclusions

- Our homology models provide structural insight into the structural determinants characterizing the substrate selectivity of CreaT.
- The presence of a  $\pi$ -helix in TM10 provides a specific packing of the binding site.
- Screening large virtual libraries would allow to validate our binding mode hypothesis.

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## RESOLUTE Research Empowerment on Solute Carriers

 Generation of a 3D model of a protein with an unknown structure ('target') based on an experimentally determined structure of a homolog protein ('template')<sup>3</sup>.

• The protocol generally includes several steps (c.f. flow chart) ranging from template selection to model validation.

• The process is iterative until a suitable model is obtained.

We modeled the CreaT transporter in the outward open and

• We included a loop refinement procedure to optimize this

